



The Possible Protective Effect of Resveratrol on Diazinon-Induced Oxidative Stress and Hepatic Injury

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Abstract

Diazinon, one of the most known organophosphate pesticides (OPs), has harmful effects on human organs. Acetylcholinesterase inhibition, oxidative stress, and inflammation are the major mechanisms of diazinon toxicity. Diazinon has several toxic effects on the liver. Resveratrol (RES) is a natural polyphenol compound with antitumor, anti-diabetic, anti-obesity, anti-oxidant, anti-aging, and anti-inflammatory effects. This compound can inhibit lipid peroxidation, protein oxidation, and DNA damage. Moreover, it can induce sirt-1, PI3K/AKT, and HO-1 pathways (negative regulators of inflammatory pathways). A large body of evidence indicated that resveratrol can attenuate liver damage by organophosphates. In this short review, we discuss the significant role of this phytoestrogen and antioxidant against the hepatotoxic effect of diazinon as an OP. With elucidation of the role of this supplement in reducing harmful effects of diazinon, it can be used as a protective agent in people at risk of adverse effects of diazinon.

Keywords: Diazinon, Oxidative stress, Inflammation, Liver, Toxicity, Resveratrol

Introduction

Pesticides are materials which are purposefully used in agriculture to increase food production. Organophosphate pesticides (OPs), as one of the most significant pesticides, have been extensively used in past decades (1). Because of the wide availability, the toxicity of OPs is a significant challenge encountered by agricultural and biological environments (2). OP poisoning is recognized as a major health problem and a common hazard in Iran (3). Diazinon (O,O-diethyl-O-[2-isopropyl-6-methyl-4-pyrimidinyl] phosphorothioate), a synthetic chemical substance, is classified as moderately hazardous (class II) organophosphorus insecticide. It has toxic effect on various organs including kidney, liver, heart, and brain, it is noted that other systems including immune and reproductive systems are affected by diazinon (4). It can be absorbed through the digestive system, skin, and respiratory tract when inhaled. Kidney has a major role in diazinon detoxification. Acetylcholinesterase suppression is a major mechanism of diazinon toxicity which leads to the accumulation of acetylcholine, prevention of signal transduction, and irreversible destructive effects on the nervous system. In addition, studies have revealed that oxidative stress could be another important mechanism involved in diazinon toxicity (5,6). Based on many studies, dose, route of exposure, physicochemical properties, and

rate of metabolism play an important role in the severity and duration of diazinon poisoning (7). Diazinon is metabolized by hepatic microsomal enzymes to hazardous metabolites such as diazoxon, hydroxyl diazoxon, and hydroxyl diazinon that have harmful effects on different organs. In hepatocytes, cytochrome P450 systems and membrane transport system of mitochondria are affected by diazinon (1). Reactive metabolites of diazinon have major role in increasing free radical production and the depletion of tissue antioxidant has a major role in hepatocellular injury (4). Reactive oxygen species (ROS) and reactive nitrogen species (RNS) formation (4), high level of malondialdehyde (MDA) (8), excessive number of abasic sites of DNA (representative of oxidative DNA damage) (9), reduction of glutathione (GSH), superoxide dismutase (SOD), catalase (CAT) (10) and vitamin C (11) are the most important procedures modulated by diazinon in oxidative stress. The apoptosis and necrosis of hepatocytes result from antioxidant depletion (12).

High levels of liver enzymes including aspartate transaminase (AST), alanine transaminase (ALT), and alkaline phosphatase (ALP), as indicators of liver damage, are reported in diazinon exposure (4). Oxidative stress damages the liver via lipid peroxidation, suppression of B-oxidation enzymes, and fatty acid accumulation in hepatocytes (13). Liver cells including Kupffer, stellate,

and endothelial cells are more sensitive to oxidative stress. Kupffer cells can produce several cytokines such as tumor necrosis factor alpha (TNF α). Concerning stellate cells, the proliferation and collagen synthesis of hepatic stellate cells are triggered by lipid peroxidation caused by oxidative stress. Additionally, oxidative stress alters metabolic pathways that control normal biological functions (14).

Organophosphates cause histoarchitectural disturbances in the liver which lead to disturbances in metabolic pathways of lipids, carbohydrates, and proteins. Previous studies indicated that diazinon increases the activity of gluconeogenesis enzymes including glycogen phosphorylase and phosphoenolpyruvate carboxykinase. Moreover, diazinon can alter liver protein metabolism (decrease in total protein and albumin) (15).

Various studies confirmed that hepatotoxicity induced by diazinon is related to inflammation (16). Diazinon increased hepatic expression and serum level of TNF- α (17). OP exposure and the high level of pro-inflammatory cytokines have a close association (16,18).

Diazinon can activate apoptotic pathways in the liver via caspase9 and caspase3 activation, increase in Bax/Bcl2 ratio, suppression of endoplasmic chaperone with anti-apoptotic properties (19,20).

It seems that implementing the supplementation strategy to ameliorate the toxic effect of diazinon may provide public health benefits. It is suggested that various substances stop the harmful effects of diazinon on the liver (21). Many studies investigated bioactive compounds extracted from plants with potential therapeutic effects (22). Plants produce low-molecular-weight secondary metabolites which have important roles in the defence system against infections or stress (23). Among them, resveratrol (RES), as a type of polyphenolic compound and phytoalexin, caught remarkable attention. Several plant species, such as peanuts, pistachios, berries, grapes, and nuts can produce RES (24). Red grape is a rich source of RES, it has been estimated that fresh grape skin contains about 50–100 g of resveratrol/g (25). Adverse conditions such as stress, injury, UV irradiation, and fungal infection can induce ROS generation (26). Pharmacological studies on RES reported anti-oxidant, anti-inflammatory, anti-cancer, anti-aging, anti-obesity, anti-diabetic, cardioprotective, neuroprotective, and anti-microbial properties of RES (27). Furthermore, health benefits of RES were indicated in treating several hepatic disorders including acetaminophen, ethanol, carbon tetrachloride, atherogenic diet induced hepatotoxicity and ischaemia/reperfusion-induced damage of the liver (28).

RES treatment can decrease thiobarbituric acid reactive substances level, increase SOD and CAT activities and inhibit activation of nuclear factor NF- κ B (as a ROS sensitive transcription factor) (29-32). Various

investigations confirmed that RES significantly suppresses cell membrane lipid peroxidation, protein oxidation, and DNA damage because it can directly scavenge different free radicals (33,34). RES is an activator of sirtuin-1 (SIRT-1) (35) which has a role in cell survival (via decreasing oxidative stress and improving forkhead box O transcription factors (36). SIRT-1 has an important function in modulating lipid metabolism, oxidative stress, and inflammation in the liver (37).

Anti-inflammatory effects of RES are indicated by direct inhibition of cyclooxygenase-2 activity (38) and reduction of gene expression of inflammatory mediators such as inducible interleukine-6 (IL-6) and interleukine-1 beta (IL-1 β) (39).

RES decreases the expression of cytochrome c, Bax, and caspase 3 and restored anti-apoptotic markers to control level (40). RES has genoprotective effect on genotoxicity induced by permethrin (an insecticide) in cultured human lymphocytes (41).

RES plays an important role in metabolic pathways. In vivo studies showed that RES can increase and decrease hepatic expression of enzymes involved in lipolysis and lipogenesis, respectively (42). In addition, RES improves glucose metabolism by increasing insulin receptor substrate 1, glucose transporter type4, peroxisome proliferator-activated receptor α and γ (43).

The hepatoprotective effects of resveratrol against fibrosis (44) and oxidative stress (45) are reported. RES can significantly decrease the expression of fibrosis-related genes including transforming growth factor beta1, collagen type-1, α -smooth muscle actin, and hydroxyproline in rat with dimethylnitrosamine induced liver fibrosis (46). The protective effect of RES against fibrogenesis is mediated by its polyphenol capacity (47).

RES treatment significantly decreased liver damage induced by CCl₄ via suppression of lipid peroxidation, improvement of hepatic GSH depletion (following CCl₄ administration), enhancement of GST activity, and nodule growth inhibition (48).

RES has beneficial effect on hepatic injury after hemorrhage via up-regulation of PI3K/AKT and HO-1 (regulators of various crucial events in the inflammatory response), and modulation of inflammatory cytokines (49).

The ameliorating effect of RES in OPs toxicity was reported. In vitro studies presented that RES can attenuate neurotoxic effects in PC12 cells via suppression of ROS production and enhancement of enzymatic antioxidant systems.

RES reduced the expression of apoptotic markers and restored the expression of anti-apoptotic markers, thereby preserving the PCL2 cells from programmed cell death (40). RES can boost the catalytic activity of Cyp2d22/

CYP2D6 in mice treated with maneb (MB), a fungicide, and paraquat (PQ), a herbicide. These xenobiotic-metabolizing enzymes have protective effects against parkinsonism induced by MB and PQ (50).

Animal model indicated that RES could inhibit the inflammatory response, oxidative stress, and apoptosis induced by paraquat (one of the world's most widely used herbicide). RES significantly declined serum AST and ALT, MDA, and hepatic TNF- α levels and the expression of apoptosis markers (including p53, Bax). In addition, this compound increased anti-apoptotic gene (BCL-2) expression and liver GSH level. Histopathological studies of the liver confirmed these results (51).

The anti-oxidative effect of resveratrol has been indicated in rats treated with glyphosate-based herbicides (GBH) via reducing MDA levels and increasing GSH levels. Moreover, RES minimized liver injury induced by GBH via improvement in biochemical markers such as AST, ALT, and ALP. These changes were reflected in the histopathological assessment of the liver (52).

Malathion is another organophosphate pesticide which significantly increases the level of liver enzymes including ALT, AST, and ALP and damages hepatocytes. In vivo studies showed that RES improved the activity of liver antioxidant enzymes and decreased lipid peroxidation and liver enzymes. Additionally, RES decreased serum nitric oxide level and lipid peroxidation in malathion-treated rats compared to the control group. RES suppresses liver damage by malathion via oxidative stress inhibition (47). RES administration can moderate the genotoxic effect of malathion in the liver by decreasing 8-hydroxyguanosine concentration. RES can improve malathion-induced histopathological changes in the liver (53). Moreover, RES has protective effects on renal damage induced by malathion. This antioxidant can reduce kidney MDA, blood urea nitrogen, nitric oxide levels and improve renal TAC level. Histopathological findings confirm these beneficial effects (54).

Figure 1 shows a summary of the antioxidant effects of RES on hepatotoxicity induced by diazinon.

Conclusion

Pesticides continue to be a substantial component in modern agriculture to increase the agricultural yield. The use of pesticides in agriculture is inevitable. The application of compounds with various anti-inflammatory, antioxidant, and hepato-protective effects such as RES can decrease the harmful effects of pesticides. In this article, we introduce a critical review of the potential preventive and therapeutic role of RES in liver toxicity induced by diazinon. Regarding the protective effects of RES, it may offer an opportunity to overcome oxidative stress and hepatocellular damage induced by diazinon. Additionally, it may be a novel choice for protection in people at risk of

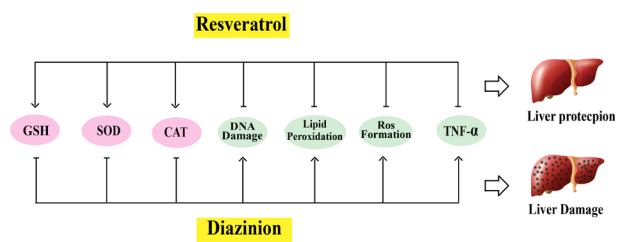


Figure 1. The Possible Effect of Resveratrol on Oxidative Stress and Inflammation Induced by Diazinon in Liver. GSH: glutathione; SOD: superoxide dismutase; CAT: catalase; ROS: reactive oxygen species; TNF- α : tumor necrosis factor α ; ↓: Inhibitory effect; ↑: Stimulatory effect.

adverse effects of diazinon.

Conflict of Interest Disclosures

The authors declare no conflicts of interest.

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References

- Gokcimen A, Gulle K, Demirin H, Bayram D, Kocak A, Altuntas I. Effects of diazinon at different doses on rat liver and pancreas tissues. *Pestic Biochem Physiol.* 2007;87(2):103-8. doi: [10.1016/j.pestbp.2006.06.011](https://doi.org/10.1016/j.pestbp.2006.06.011).
- Shah MD, Iqbal M. Diazinon-induced oxidative stress and renal dysfunction in rats. *Food Chem Toxicol.* 2010;48(12):3345-53. doi: [10.1016/j.fct.2010.09.003](https://doi.org/10.1016/j.fct.2010.09.003).
- Alinejad S, Zamani N, Abdollahi M, Mehrpour O. A narrative review of acute adult poisoning in Iran. *Iran J Med Sci.* 2017;42(4):327-46.
- Beydilli H, Yilmaz N, Cetin ES, Topal Y, Celik OI, Sahin C, et al. Evaluation of the protective effect of silibinin against diazinon induced hepatotoxicity and free-radical damage in rat liver. *Iran Red Crescent Med J.* 2015;17(4):e25310. doi: [10.5812/ircmj.17\(4\)2015.25310](https://doi.org/10.5812/ircmj.17(4)2015.25310).
- Abu-Qare AW, Abou-Donia MB. Inhibition and recovery of maternal and fetal cholinesterase enzyme activity following a single cutaneous dose of methyl parathion and diazinon, alone and in combination, in pregnant rats. *J Appl Toxicol.* 2001;21(4):307-16. doi: [10.1002/jat.761](https://doi.org/10.1002/jat.761).
- Sams C, Cocker J, Lennard MS. 544 Metabolism of chlorpyrifos and diazinon by human liver microsomes. *Toxicol Lett.* 2003;144 Suppl 1:s146. doi: [10.1016/s0378-4274\(03\)90543-1](https://doi.org/10.1016/s0378-4274(03)90543-1).
- Karalliedde LD, Edwards P, Marrs TC. Variables influencing the toxic response to organophosphates in humans. *Food Chem Toxicol.* 2003;41(1):1-13. doi: [10.1016/s0278-6915\(02\)00232-6](https://doi.org/10.1016/s0278-6915(02)00232-6).
- El-Shenawy NS, El-Salmy F, Al-Eisa RA, El-Ahmary B. Amelioratory effect of vitamin E on organophosphorus insecticide diazinon-induced oxidative stress in mice liver. *Pestic Biochem Physiol.* 2010;96(2):101-7. doi: [10.1016/j.pestbp.2009.09.008](https://doi.org/10.1016/j.pestbp.2009.09.008).
- Tsitsimpikou C, Tzatzarakis M, Fragkiadaki P, Kovatsi L, Stivaktakis P, Kalogeraki A, et al. Histopathological lesions, oxidative stress and genotoxic effects in liver and kidneys following long term exposure of rabbits to diazinon and propoxur. *Toxicology.* 2013;307:109-14. doi: [46](https://doi.org/10.1016/j.

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- tox.2012.11.002.
10. Messarah M, Amamra W, Boumendjel A, Barkat L, Bouasla I, Abdennour C, et al. Ameliorating effects of curcumin and vitamin E on diazinon-induced oxidative damage in rat liver and erythrocytes. *Toxicol Ind Health*. 2013;29(1):77-88. doi: [10.1177/0748233712446726](https://doi.org/10.1177/0748233712446726).
 11. Abdel-Daim MM. Synergistic protective role of ceftriaxone and ascorbic acid against subacute diazinon-induced nephrotoxicity in rats. *Cytotechnology*. 2016;68(2):279-89. doi: [10.1007/s10616-014-9779-z](https://doi.org/10.1007/s10616-014-9779-z).
 12. Li S, Tan HY, Wang N, Zhang ZJ, Lao L, Wong CW, et al. The role of oxidative stress and antioxidants in liver diseases. *Int J Mol Sci*. 2015;16(11):26087-124. doi: [10.3390/ijms161125942](https://doi.org/10.3390/ijms161125942).
 13. Reddy JK, Rao MS. Lipid metabolism and liver inflammation. II. Fatty liver disease and fatty acid oxidation. *Am J Physiol Gastrointest Liver Physiol*. 2006;290(5):G852-8. doi: [10.1152/ajpgi.00521.2005](https://doi.org/10.1152/ajpgi.00521.2005).
 14. Le Lay S, Simard G, Martinez MC, Andriantsitohaina R. Oxidative stress and metabolic pathologies: from an adipocentric point of view. *Oxid Med Cell Longev*. 2014;2014:908539. doi: [10.1155/2014/908539](https://doi.org/10.1155/2014/908539).
 15. Teimouri F, Amirkabirian N, Esmaily H, Mohammadirad A, Aliahmadi A, Abdollahi M. Alteration of hepatic cells glucose metabolism as a non-cholinergic detoxication mechanism in counteracting diazinon-induced oxidative stress. *Hum Exp Toxicol*. 2006;25(12):697-703. doi: [10.1177/0960327106075064](https://doi.org/10.1177/0960327106075064).
 16. Timcheh Hariri A, Moallem SA, Mahmoudi M, Memar B, Hosseinzadeh H. Sub-acute effects of diazinon on biochemical indices and specific biomarkers in rats: protective effects of crocin and safranal. *Food Chem Toxicol*. 2010;48(10):2803-8. doi: [10.1016/j.fct.2010.07.010](https://doi.org/10.1016/j.fct.2010.07.010).
 17. Ahmadi-Naji R, Heidarian E, Ghatreh-Samani K. Evaluation of the effects of the hydroalcoholic extract of *Terminalia chebula* fruits on diazinon-induced liver toxicity and oxidative stress in rats. *Avicenna J Phytomed*. 2017;7(5):454-66.
 18. Gangemi S, Gofita E, Costa C, Teodoro M, Briguglio G, Nikitovic D, et al. Occupational and environmental exposure to pesticides and cytokine pathways in chronic diseases (Review). *Int J Mol Med*. 2016;38(4):1012-20. doi: [10.3892/ijmm.2016.2728](https://doi.org/10.3892/ijmm.2016.2728).
 19. Karami-Mohajeri S, Ahmadipour A, Rahimi HR, Abdollahi M. Adverse effects of organophosphorus pesticides on the liver: a brief summary of four decades of research. *Arh Hig Rada Toksikol*. 2017;68(4):261-75. doi: [10.1515/aiht-2017-68-2989](https://doi.org/10.1515/aiht-2017-68-2989).
 20. Lari P, Rashedinia M, Abnous K, Hosseinzadeh H. Crocin improves lipid dysregulation in subacute diazinon exposure through ERK1/2 pathway in rat liver. *Drug Res (Stuttg)*. 2014;64(6):301-5. doi: [10.1055/s-0033-1357196](https://doi.org/10.1055/s-0033-1357196).
 21. Abdel-Daim MM, Taha R, Ghazy EW, El-Sayed YS. Synergistic ameliorative effects of sesame oil and alpha-lipoic acid against subacute diazinon toxicity in rats: hematological, biochemical, and antioxidant studies. *Can J Physiol Pharmacol*. 2016;94(1):81-8. doi: [10.1139/cjpp-2015-0131](https://doi.org/10.1139/cjpp-2015-0131).
 22. Bortolotti C, Kunit T, Moder A, Hufnagl C, Schmidt S, Hartl A, et al. The phytoestrogen resveratrol induces apoptosis in INS-1E rat insulinoma cells. *Cell Physiol Biochem*. 2009;23(4-6):245-54. doi: [10.1159/000218171](https://doi.org/10.1159/000218171).
 23. Borriello A, Cucciolla V, Della Ragione F, Galletti P. Dietary polyphenols: focus on resveratrol, a promising agent in the prevention of cardiovascular diseases and control of glucose homeostasis. *Nutr Metab Cardiovasc Dis*. 2010;20(8):618-25. doi: [10.1016/j.numecd.2010.07.004](https://doi.org/10.1016/j.numecd.2010.07.004).
 24. Ndiaye M, Philippe C, Mukhtar H, Ahmad N. The grape antioxidant resveratrol for skin disorders: promise, prospects, and challenges. *Arch Biochem Biophys*. 2011;508(2):164-70. doi: [10.1016/j.abb.2010.12.030](https://doi.org/10.1016/j.abb.2010.12.030).
 25. Floreani M, Napoli E, Quintieri L, Palatini P. Oral administration of trans-resveratrol to guinea pigs increases cardiac DT-diaphorase and catalase activities, and protects isolated atria from menadione toxicity. *Life Sci*. 2003;72(24):2741-50. doi: [10.1016/s0024-3205\(03\)00179-6](https://doi.org/10.1016/s0024-3205(03)00179-6).
 26. Abbasi Oshaghi E, Goodarzi MT, Higgins V, Adeli K. Role of resveratrol in the management of insulin resistance and related conditions: mechanism of action. *Crit Rev Clin Lab Sci*. 2017;54(4):267-93. doi: [10.1080/10408363.2017.1343274](https://doi.org/10.1080/10408363.2017.1343274).
 27. Rauf A, Imran M, Suleria HAR, Ahmad B, Peters DG, Mubarak MS. A comprehensive review of the health perspectives of resveratrol. *Food Funct*. 2017;8(12):4284-305. doi: [10.1039/c7fo01300k](https://doi.org/10.1039/c7fo01300k).
 28. Bishayee A, Darvesh AS, Politis T, McGory R. Resveratrol and liver disease: from bench to bedside and community. *Liver Int*. 2010;30(8):1103-14. doi: [10.1111/j.1478-3231.2010.02295.x](https://doi.org/10.1111/j.1478-3231.2010.02295.x).
 29. Leonard SS, Xia C, Jiang BH, Stinefelt B, Klandorf H, Harris GK, et al. Resveratrol scavenges reactive oxygen species and effects radical-induced cellular responses. *Biochem Biophys Res Commun*. 2003;309(4):1017-26. doi: [10.1016/j.bbrc.2003.08.105](https://doi.org/10.1016/j.bbrc.2003.08.105).
 30. Schmatz R, Perreira LB, Stefanello N, Mazzanti C, Spanevello R, Gutierrez J, et al. Effects of resveratrol on biomarkers of oxidative stress and on the activity of delta aminolevulinic acid dehydratase in liver and kidney of streptozotocin-induced diabetic rats. *Biochimie*. 2012;94(2):374-83. doi: [10.1016/j.biochi.2011.08.005](https://doi.org/10.1016/j.biochi.2011.08.005).
 31. Asadi S, Rahimi Z, Saidijam M, Shabab N, Goodarzi MT. Effects of resveratrol on FOXO1 and FOXO3a genes expression in adipose tissue, serum insulin, insulin resistance and serum SOD activity in type 2 diabetic rats. *Int J Mol Cell Med*. 2018;7(3):176-84. doi: [10.22088/ijmcm.bums.7.3.176](https://doi.org/10.22088/ijmcm.bums.7.3.176).
 32. Asadi S, Moradi MN, Khyripour N, Goodarzi MT, Mahmoodi M. Resveratrol attenuates copper and zinc homeostasis and ameliorates oxidative stress in type 2 diabetic rats. *Biol Trace Elem Res*. 2017;177(1):132-8. doi: [10.1007/s12011-016-0861-6](https://doi.org/10.1007/s12011-016-0861-6).
 33. Sengottuvelan M, Deeptha K, Nalini N. Resveratrol ameliorates DNA damage, prooxidant and antioxidant imbalance in 1,2-dimethylhydrazine induced rat colon carcinogenesis. *Chem Biol Interact*. 2009;181(2):193-201. doi: [10.1016/j.cbi.2009.06.004](https://doi.org/10.1016/j.cbi.2009.06.004).
 34. Xiao NN. Effects of resveratrol supplementation on oxidative damage and lipid peroxidation induced by strenuous exercise in rats. *Biomol Ther (Seoul)*. 2015;23(4):374-8. doi: [10.4062/biomolther.2015.015](https://doi.org/10.4062/biomolther.2015.015).
 35. Borra MT, Smith BC, Denu JM. Mechanism of human SIRT1 activation by resveratrol. *J Biol Chem*. 2005;280(17):17187-95. doi: [10.1074/jbc.M501250200](https://doi.org/10.1074/jbc.M501250200).
 36. Pan H, Finkel T. Key proteins and pathways that regulate lifespan. *J Biol Chem*. 2017;292(16):6452-60. doi: [10.1074/jbc.R116.771915](https://doi.org/10.1074/jbc.R116.771915).
 37. Ding RB, Bao J, Deng CX. Emerging roles of SIRT1 in fatty liver diseases. *Int J Biol Sci*. 2017;13(7):852-67. doi: [10.7150/ijbs.19370](https://doi.org/10.7150/ijbs.19370).
 38. Subbaramaiah K, Chung WJ, Michaluart P, Telang N, Tanabe T, Inoue H, et al. Resveratrol inhibits cyclooxygenase-2 transcription and activity in phorbol ester-treated human mammary epithelial cells. *J Biol Chem*. 1998;273(34):21875-82. doi: [10.1074/jbc.273.34.21875](https://doi.org/10.1074/jbc.273.34.21875).
 39. Limagne E, Lançon A, Delmas D, Cherkaoui-Malki M, Latruffe N. Resveratrol Interferes with IL1-β-Induced Pro-Inflammatory Paracrine Interaction between primary chondrocytes and macrophages. *nutrients*. 2016;8(5). doi: [10.3390/nu8050280](https://doi.org/10.3390/nu8050280).
 40. Kumar V, Tripathi VK, Singh AK, Lohani M, Kuddus M. Trans-resveratrol restores the damages induced by organophosphate pesticide-monocrotophos in neuronal cells. *Toxicol Int*.

- 2013;20(1):48-55. doi: [10.4103/0971-6580.111571](https://doi.org/10.4103/0971-6580.111571).
41. Turkez H, Aydin E. The genoprotective activity of resveratrol on permethrin-induced genotoxic damage in cultured human lymphocytes. *Braz Arch Biol Technol*. 2013;56(3):405-11. doi: [10.1590/s1516-89132013000300008](https://doi.org/10.1590/s1516-89132013000300008).
 42. Zhang D, Yan Y, Tian H, Jiang G, Li X, Liu W. Resveratrol supplementation improves lipid and glucose metabolism in high-fat diet-fed blunt snout bream. *Fish Physiol Biochem*. 2018;44(1):163-73. doi: [10.1007/s10695-017-0421-9](https://doi.org/10.1007/s10695-017-0421-9).
 43. Faghihzadeh F, Hekmatdoost A, Adibi P. Resveratrol and liver: a systematic review. *J Res Med Sci*. 2015;20(8):797-810. doi: [10.4103/1735-1995.168405](https://doi.org/10.4103/1735-1995.168405).
 44. Mohseni R, Arab Sadeghabadi Z, Goodarzi MT, Karimi J. Co-administration of resveratrol and beta-aminopropionitrile attenuates liver fibrosis development via targeting lysyl oxidase in CCl₄-induced liver fibrosis in rats. *Immunopharmacol Immunotoxicol*. 2019;41(6):644-51. doi: [10.1080/08923973.2019.1688829](https://doi.org/10.1080/08923973.2019.1688829).
 45. Khazaei M, Karimi J, Sheikh N, Goodarzi MT, Saidijam M, Khodadadi I, et al. Effects of resveratrol on receptor for advanced glycation end products (RAGE) expression and oxidative stress in the liver of rats with type 2 diabetes. *Phytother Res*. 2016;30(1):66-71. doi: [10.1002/ptr.5501](https://doi.org/10.1002/ptr.5501).
 46. Hong SW, Jung KH, Zheng HM, Lee HS, Suh JK, Park IS, et al. The protective effect of resveratrol on dimethylnitrosamine-induced liver fibrosis in rats. *Arch Pharm Res*. 2010;33(4):601-9. doi: [10.1007/s12272-010-0415-y](https://doi.org/10.1007/s12272-010-0415-y).
 47. Jalili C, Farzaei MH, Roshankhah S, Salahshoor MR. Resveratrol attenuates malathion-induced liver damage by reducing oxidative stress. *J Lab Physicians*. 2019;11(3):212-9. doi: [10.4103/jlp.jlp_43_19](https://doi.org/10.4103/jlp.jlp_43_19).
 48. Roy S, Sannigrahi S, Majumdar S, Ghosh B, Sarkar B. Resveratrol regulates antioxidant status, inhibits cytokine expression and restricts apoptosis in carbon tetrachloride induced rat hepatic injury. *Oxid Med Cell Longev*. 2011;2011:703676. doi: [10.1155/2011/703676](https://doi.org/10.1155/2011/703676).
 49. Yu HP, Yang SC, Lau YT, Hwang TL. Role of Akt-dependent up-regulation of hemoxygenase-1 in resveratrol-mediated attenuation of hepatic injury after trauma hemorrhage. *Surgery*. 2010;148(1):103-9. doi: [10.1016/j.surg.2009.12.008](https://doi.org/10.1016/j.surg.2009.12.008).
 50. Srivastava G, Dixit A, Yadav S, Patel DK, Prakash O, Singh MP. Resveratrol potentiates cytochrome P450 2 d22-mediated neuroprotection in maneb- and paraquat-induced parkinsonism in the mouse. *Free Radic Biol Med*. 2012;52(8):1294-306. doi: [10.1016/j.freeradbiomed.2012.02.005](https://doi.org/10.1016/j.freeradbiomed.2012.02.005).
 51. El-Boghdady NA, Abdeltawab NF, Nooh MM. Resveratrol and montelukast alleviate paraquat-induced hepatic injury in mice: modulation of oxidative stress, inflammation, and apoptosis. *Oxid Med Cell Longev*. 2017;2017:9396425. doi: [10.1155/2017/9396425](https://doi.org/10.1155/2017/9396425).
 52. Turkmen R, Birdane YO, Demirel HH, Kabu M, Ince S. Protective effects of resveratrol on biomarkers of oxidative stress, biochemical and histopathological changes induced by sub-chronic oral glyphosate-based herbicide in rats. *Toxicol Res (Camb)*. 2019;8(2):238-45. doi: [10.1039/c8tx00287h](https://doi.org/10.1039/c8tx00287h).
 53. Akbel E, Arslan-Acaroz D, Demirel HH, Kucukkurt I, Ince S. The subchronic exposure to malathion, an organophosphate pesticide, causes lipid peroxidation, oxidative stress, and tissue damage in rats: the protective role of resveratrol. *Toxicol Res (Camb)*. 2018;7(3):503-12. doi: [10.1039/c8tx00030a](https://doi.org/10.1039/c8tx00030a).
 54. Jalili C, Roshankhah S, Moradi Y, Salahshoor MR. Resveratrol attenuates malathion-induced renal damage by declining oxidative stress in rats. *Int J Pharm Investig*. 2018;8(4):192-9.